

DITERPENES FROM *SIDERITIS INFERNALIS*

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Abstract—Five new *ent*-kaurene diterpenes, episinferral, sinferral, sinfernol, epoxysinferral and canditriol, and the known one, candol B, candicandiol and candidiol, have been isolated from the aerial parts of *Sideritis infernalis*.

INTRODUCTION

The phytochemical study of the genus *Sideritis* (Labiatae), endemic to the Canary Islands, has been a subject of interest to us during the past 15 years [1, 2]. We have now investigated the aerial parts of *Sideritis infernalis*, a species endemic to the island of Tenerife, from which we have obtained five new diterpenes: episinferral (*ent*-7 α ,18-dihydroxykaur-15-en-17-al, 1), sinferral (*ent*-7 β ,18-dihydroxykaur-15-en-17-al, 5), sinfernol (*ent*-7 β ,17,18-trihydroxykaur-15-ene, 7), epoxysinferral (*ent*-15 β ,16 β -epoxy-7 β ,17,18-trihydroxykaurane, 18) and canditriol (*ent*-7 β ,15 β ,18-trihydroxykaur-16-ene, 21).

RESULTS AND DISCUSSION

The structure 1 was assigned to episinferral, the least polar new substance obtained from *S. infernalis*. This compound was isolated as its diacetate 2 by acetylation and chromatography of fractions obtained from the main chromatography. Its ¹H NMR spectrum showed the signals of two methyls, two acetates, the two hydrogens of an acetylated primary alcohol (δ 3.68, s) the proton geminal to an acetylated secondary alcohol (δ 4.81, br s), an aldehydic proton (δ 9.71, s) and the hydrogen of a double bond conjugated with the carbonyl of the aldehyde (δ 6.66, s). On the basis of these data and comparison with those of epicandicandiol diacetate (3) [3], we assigned this compound a kaurene skeleton with the two acetylated hydroxyls at C-7 β and C-18. The rest of the molecule must be formed with the aldehyde at C-17 and the double bond at C-15, C-16. The ¹³C NMR spectral data (Table 1) confirmed the structure of episinferral diacetate (2).

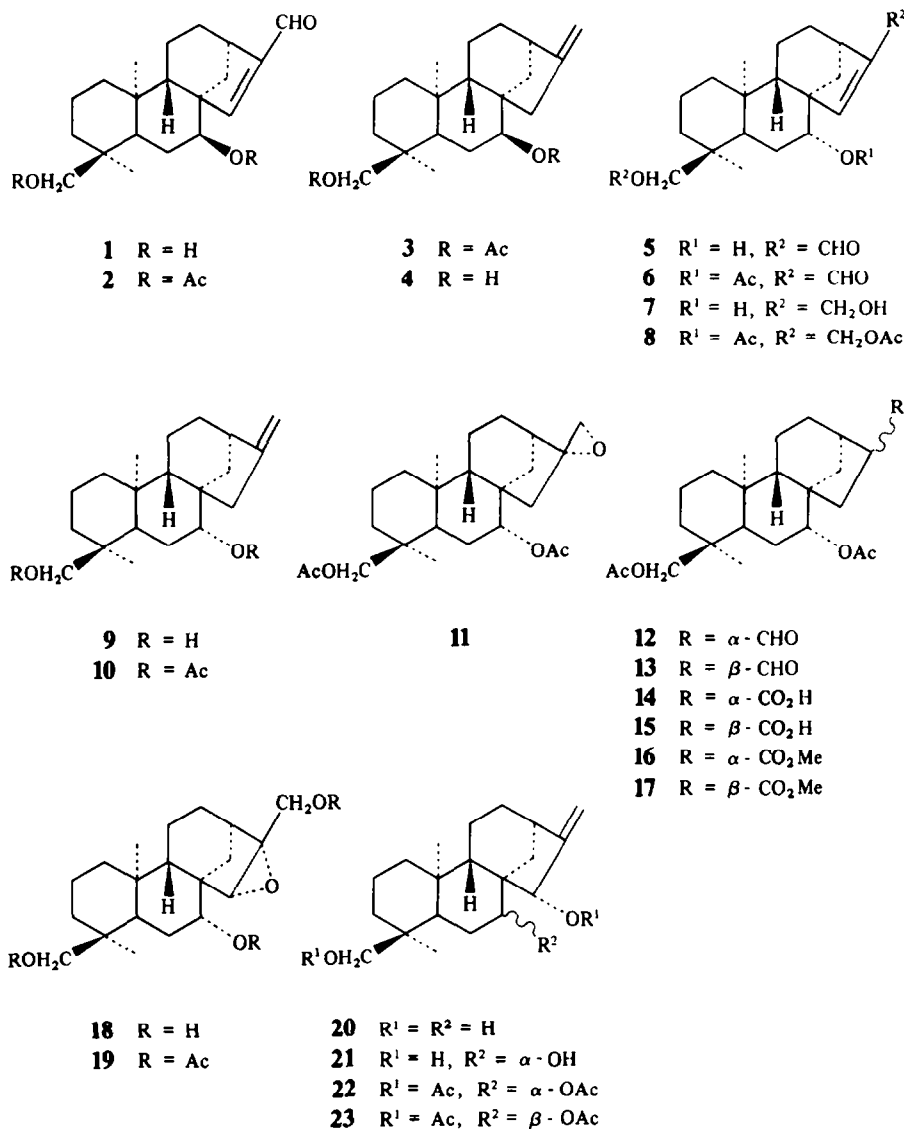
The second new diterpene was named sinferral (5). The ¹H NMR spectrum of its diacetate 6 was similar to that of 2, but the resonance of the proton geminal to the acetylated secondary alcohol was different in both compounds, now being equatorial in 6 (4.96, dd, *J* = 11 and 4 Hz). Although the ¹³C NMR spectrum confirmed the structure of 6, the new compound sinferral (5) was related to candicandiol (9) [4, 5] in the following way. The diacetate of candicandiol (10) was epoxidized with *m*-chloroperbenzoic acid giving the epoxide 11. Treatment of this compound with boron trifluoride etherate in benzene afforded a mixture of the aldehydes 12 and 13 in

the proportion 2:1 (¹H NMR: δ 9.81 and 9.60). This mixture was easily oxidized by air forming the acids 14 and 15, which were separated by chromatography and methylated with diazomethane to give the corresponding methyl esters 16 and 17. These two compounds were identical with those obtained by hydrogenation of sinferral diacetate (6), followed by air oxidation and methylation. The configurations at C-16 of the two acids, 14 and 15, were assigned in accordance with the magnitude of the H-13, H-16 coupling patterns in the ¹H NMR spectra of their methyl esters 16 and 17.

The third new compound, sinfernol (7), was a triol related to compound 5. In its ¹H NMR spectrum the aldehydic proton had disappeared; being replaced now by the two doublets characteristic of a hydroxymethylene group, and the hydrogen of the double bond was now at a higher field than seen in 5. In accordance with these data, the structure 7 was assigned to this new compound. Reduction of sinferral (5) with sodium borohydride in the presence of cerium trichloride [6] afforded 7, identical with the natural compound sinfernol.

Another new diterpene isolated from this species was epoxysinferral (18). This compound was obtained in the form of the triacetate (19) by acetylation of some fractions of the extract that contained 18. The ¹H NMR spectrum of 19 was also very similar to that of sinfernol triacetate (8), but it lacked the vinyl proton signal, a new one appearing that originated from the hydrogen geminal to an epoxidic function. Epoxidation of sinfernol triacetate (8) with *m*-chloroperbenzoic acid afforded the epoxide 19, identical with the acetylated natural product. The α -configuration for the oxirane ring was given because it is known that in this kind of compound oxidation occurs on this face.

Finally, the most polar new compound was named canditriol and its structure determined as 21 on the basis of the following considerations. High resolution mass spectrometry was in accordance with the formula C₂₀H₃₂O₃. Its ¹H NMR spectrum displayed signals of a primary alcohol group, which must be equatorial as inferred from the chemical shift of the hydroxymethylene hydrogens in 21 and its acetate 22 [7]. Other signals observed in this spectrum were two methyl groups, an exocyclic double bond and two protons geminal to hydroxylic functions. One of these alcoholic groups must



be at C-7α, because in comparison with candicandiol its geminal proton has the same form of resonance. The other must be at C-15 because its geminal hydrogen appears in the spectrum as a singlet at δ4.06. The α-configuration was given to the C-15 alcohol on consideration that this plant also contains candiol (20), a diterpene with a 15α-hydroxyl function. Oxidation of candicandiol (9) with selenium dioxide afforded a compound (21) identical with the natural substance. This reaction also confirmed the stereochemistry of the alcohol group at C-15, because it is known that in this reaction the hydroxyl formed has the α-stereochemistry [8].

Sideritriol, epoxysideritriol and eubotriol, the C-7 epimers of sinfernol (7), epoxysinfernol (18) and canditriol (21) respectively, have been isolated from other species of *Sideritis* genus [9–11]. We have now prepared eubotriol by selenium dioxide oxidation of epicandicandiol (4), and the ¹³C NMR spectral data of its triacetate (23) is shown in Table 1.

We have also isolated from this species the known diterpenes candol B (18-hydroxy-*ent*-kaur-16-ene) [12], candiol (20) [3] and candicandiol (9) [4, 5].

EXPERIMENTAL

Mps: uncorr.; NMR: CDCl₃, 200 MHz, unless indicated otherwise; MS: 70 eV (probe); column and dry column chromatography: silica gel 0.063–0.2 mm.

The air-dried aerial part of the plant, *Sideritis infernalis* (1.7 kg), collected at Barranco del Infierno (Adeje, Tenerife) in May, was used for this work. A voucher specimen has been deposited at the Herbarium of the Instituto Canario de Investigaciones Agrarias (ORT 28643).

A general description of the procedure to isolate the terpenoid substances of species of the genus *Sideritis* has been published previously [2]. In this way candol B (15 mg), candiol (20, 3.5 g), candicandiol (9, 5.9 g), a mixture of episinferral (1) and sinfernol (5) (60 mg), sinfernol (5, 120 mg), sinfernol (7, 80 mg), a mixture of sinfernol (7) and epoxysinfernol (18) (130 mg), and canditriol (21, 160 mg) were obtained. The mixture of the aldehydes 1 and 5 was acetylated and chromatographed to give pure compounds 2 (20 mg) and 6 (25 mg). In the same way the mixture of 7 and 18 was resolved as its acetates 8 (40 mg) and 19 (70 mg).

Episinferral diacetate (2). Mp 125–127°. [M]⁺ at m/z 402.2402. C₂₄H₃₄O₅ requires 402.2404; ¹H NMR: δ0.80 and 1.08 (each 3H,

Table 1. ^{13}C NMR spectral data (CDCl_3 , 50.32 MHz)

Carbon	2	3*	6	8	10	19	22	23
1	39.68	39.5	39.73	39.09	39.68	39.77	39.53	39.56
2	17.76	17.6	17.75	17.85	17.72	17.70	17.73	17.78
3	35.66	35.5	35.70	35.92†	35.73	35.65	35.72	35.46
4	36.29	36.1	36.48	36.48	36.23	36.50	36.25	36.21
5	41.55	41.7	46.35†	46.73	46.55	46.79	45.59	41.57
6	23.89	24.8	25.29	25.60	25.56	26.63	26.35	24.20
7	76.00	79.3	76.53	75.99	76.40	74.59	73.11	74.68
8	53.65	46.8	55.61	53.84	48.19	47.49	50.76	49.79
9	43.10	51.2	46.69†	47.66	55.21	49.25	54.06	48.92
10	39.68	38.9	39.24	40.46	39.10	38.88	38.99	39.43
11	18.14	17.6	18.33	18.48	17.89	18.05	17.99	17.78
12	25.14	33.2	25.12	25.34	33.30	25.34	32.64	33.32
13	38.07	43.6	36.79	39.80	42.81	34.94	41.71	42.27
14	41.39	38.2	35.31	35.84†	31.84	24.64	30.94	35.38
15	155.63	45.1	158.47	135.32	43.00	63.50	82.14	80.49
16	149.38	154.1	149.25	142.04	154.38	61.20	154.85	154.95
17	189.44	103.7	189.15	62.33	103.32	62.04	109.68	110.60
18	72.53	72.4	72.31	72.42	72.21	72.35	72.20	72.52
19	17.39	17.3	17.56	17.60	17.53	17.63	17.43	17.60
20	18.03	17.8	18.29	18.28	18.06	17.92	17.86	18.02

*Taken from ref. [13].

†These values can be interchanged.

s), 2.04 and 2.11 (each 3H, s), 3.68 (2H, s, H-18), 4.81 (1H, br s, H-7), 6.66 (1H, s, H-15), 9.71 (1H, s, H-17); EIMS m/z (rel. int.): 402 $[\text{M}]^+$ (0.2), 360 (3), 342 (8), 300 (4), 298 (5), 285 (17), 282 (18), 269 (43), 267 (19), 254 (7), 253 (9), 251 (8), 239 (6), 225 (9).

Sinferral (5). Obtained as a gum; ^1H NMR: δ 0.72 and 1.07 (each 3H, s), 3.06 and 3.37 (each 1H, d , J = 10 Hz, H-18), 3.85 (1H, br s, H-7), 6.70 (1H, s, H-15), 9.71 (1H, s, H-17); EIMS m/z (rel. int.): 318 $[\text{M}]^+$ (0.5), 303 (0.7), 300 (1), 287 (29), 275 (4), 269 (6), 257 (3), 161 (4), 149 (17), 145 (4), 135 (6), 131 (4), 123 (13), 117 (5), 109 (16). Sinferral diacetate (6), a gum $[\text{M}]^+$ at m/z 402.2355, $\text{C}_{24}\text{H}_{34}\text{O}_5$ requires 402.2406; ^1H NMR: δ 0.77 and 1.06 (each 3H, s), 1.95 and 2.03 (each 3H, s), 1.95 and 2.03 (each 3H, s), 3.55 and 3.77 (each 1H, d , J = 10 Hz, H-18), 4.96 (1H, dd , J = 11 and 4 Hz, H-7), 6.62 (1H, s, H-15), 9.64 (1H, s, H-17); EIMS m/z (rel. int.): 402 $[\text{M}]^+$ (0.6), 360 (7), 342 (5), 300 (4), 282 (25), 270 (12), 269 (52), 267 (17), 254 (9), 225 (7).

Sinferral (7). Mp 225–227°, $[\text{M}]^+$ at m/z 320.2312, $\text{C}_{20}\text{H}_{32}\text{O}_3$ requires 320.2351; ^1H NMR (C_6D_6): δ 0.84 and 1.11 (each 3H, s), 3.32 and 3.69 (each 1H, d , J = 10 Hz, H-18), 4.12 (1H, br s, H-7), 4.49 (2H, s, H-17), 5.81 (1H, s, H-15); EIMS m/z (rel. int.): 320 $[\text{M}]^+$ (1), 302 (12), 287 (22), 271 (18), 253 (7), 209 (7), 180 (6), 162 (13), 161 (10), 157 (6), 145 (11), 143 (6), 131 (12). Sinferral triacetate (8), $[\text{M} - \text{C}_2\text{H}_2\text{O}]^+$ at m/z 404.2539, $\text{C}_{24}\text{H}_{36}\text{O}_5$ requires 404.2561; ^1H NMR: δ 0.79 and 1.06 (each 3H, s), 1.99, 2.05 and 2.07 (each 3H, s), 3.58 and 3.80 (each 1H, d , J = 11 Hz, H-18), 4.53 and 4.63 (each 1H, d , J = 14 Hz, H-17), 4.89 (1H, dd , J = 11 and 5 Hz, H-7), 5.44 (1H, s, H-15); EIMS m/z (rel. int.): 404 $[\text{M} - \text{C}_2\text{H}_2\text{O}]^+$ (2), 386 (18), 345 (21), 344 (78), 327 (12), 326 (43), 311 (11), 284 (22), 266 (49), 253 (61), 251 (35), 237 (13), 225 (17), 223 (14), 197 (12).

Epoxysinferral triacetate (19). $[\text{M} - \text{C}_2\text{H}_2\text{O}]^+$ at m/z 420.2878, $\text{C}_{24}\text{H}_{36}\text{O}_6$ requires 420.2512; ^1H NMR: δ 0.78 and 1.02 (each 3H, s), 2.02 (3H, s), 2.04 (6H, s), 3.00 (1H, s, H-15), 3.59 and 3.76 (each 1H, d , J = 11 Hz, H-18), 3.98 and 4.62 (each 1H, d , J = 12 Hz, H-17), 5.01 (1H, dd , J = 11 and 5 Hz, H-7); EIMS m/z (rel. int.): 420 $[\text{M} - \text{C}_2\text{H}_2\text{O}]^+$ (1), 403 (51), 386 (3), 360 (2), 344 (6), 326 (5), 313 (4), 283 (11), 253 (9), 225 (5), 197 (4).

Canditriol (21). Mp 220–222°, $[\text{M}]^+$ at m/z 320.2393, $\text{C}_{20}\text{H}_{32}\text{O}_3$ requires 320.2352; ^1H NMR: δ 0.72 and 1.05 (each 3H, s), 3.04 and 3.45 (each 1H, d , J = 11 Hz, H-18), 3.88 (1H, dd , J = 11 and 5 Hz, H-7), 4.06 (1H, s, H-15), 5.05 and 5.13 (each 1H, s, H-17); EIMS m/z (rel. int.): 320 $[\text{M}]^+$ (15), 302 (13), 287 (8), 271 (16), 262 (21), 261 (7), 253 (7), 201 (8), 180 (5), 162 (19), 147 (20). Canditriol triacetate (22), $[\text{M} - \text{C}_2\text{H}_2\text{O}]^+$ at m/z 404.2578, $\text{C}_{24}\text{H}_{36}\text{O}_5$ requires 404.2560; ^1H NMR: δ 0.77 and 1.06 (each 3H, s), 1.96, 2.00 and 2.07 (each 3H, s), 3.60 and 3.77 (each 1H, d , J = 11 Hz, H-18), 4.81 (1H, dd , J = 11 and 5 Hz, H-7), 5.01 and 5.06 (each 1H, s, H-17), 5.41 (1H, s, H-15); EIMS m/z (rel. int.): 404 $[\text{M} - \text{C}_2\text{H}_2\text{O}]^+$ (23), 386 (10), 362 (6), 344 (71), 326 (26), 313 (15), 284 (31), 283 (22), 269 (28), 253 (36), 251 (24).

Epoxidation of compound 8. Sinferral triacetate (8, 17 mg) in CHCl_3 (2 ml) was added to a soln of *m*-chloroperbenzoic acid (10 mg) in CHCl_3 (2 ml). The mixture was left at room temp. for 90 min, washed with a satd soln of NaHCO_3 and then usual work-up and chromatography of the residue gave 19, identical with the acetylated natural substance.

Epoxidation of compound 10. Candicandiol diacetate (10, 300 mg) in CHCl_3 (8 ml) was treated as above for 8. The reaction time in this case was 15 hr. Usual work-up afforded 11 (250 mg), ^1H NMR: δ 0.77 and 1.06 (each 3H, s), 2.02 and 2.04 (each 3H, s), 2.76 and 2.84 (each 1H, d , J = 5 Hz, H-17), 3.54 and 3.78 (each 1H, d , J = 11 Hz, H-18), 4.64 (1H, dd , J = 11 and 5 Hz, H-7); EIMS m/z (rel. int.): 362 $[\text{M} - \text{C}_2\text{H}_2\text{O}]^+$ (1), 344 (5), 329 (2), 289 (37), 284 (87), 271 (94), 269 (53), 255 (24), 253 (22), 241 (16), 227 (40), 225 (11), 213 (16), 201 (11), 199 (13), 197 (13).

Preparation of compounds 16 and 17. Compound 11 (230 mg) in dry C_6H_6 (9 ml) was treated with $\text{BF}_3 \cdot \text{Et}_2\text{O}$ complex (3 drops) at 0° for 30 min. Usual work-up, evaporation of the solvent and chromatography of the residue gave a mixture of aldehydes 12 and 13 (200 mg). They were easily oxidized by air to a mixture of acids, which were separated by chromatography. Elution with C_6H_6 – EtOAc (10%) afforded 14, mp 207–209°; ^1H NMR: δ 0.78 and 1.03 (each 3H, s), 2.01 and 2.05 (each 3H, s), 3.58 and 3.79 (each 1H, d , J = 11 Hz, H-18), 4.71 (1H, dd , J = 11 and 5 Hz, H-

7). Further elution gave **15**, mp 209–211°. ¹H NMR (200 MHz): δ 0.77 and 1.03 (each 3H, s), 2.01 and 2.06 (each 3H, s), 3.55 and 3.80 (each 1H, d, *J* = 11 Hz, H-18), 4.65 (1H, m, H-7). Treatment of **14** and **15**, separately, with CH₂N₂ gave the corresponding methyl esters, **16** and **17**. *Methyl ester 16*. ¹H NMR: δ 0.79 and 1.04 (each 3H, s), 2.06 (6H, s), 2.45 (1H, br s, H-13), 2.60 (1H, dd, *J*₁ = 5, *J*₂ = 9 Hz, H-16), 3.58 and 3.79 (each 1H, d, *J* = 11 Hz, H-18), 3.62 (3H, s), 4.71 (1H, dd, *J* = 11 and 5 Hz, H-7); EIMS *m/z* (rel. int.): 403 [M – MeO]⁺, 392 (2), 374 (3), 359 (2), 332 (4), 314 (79), 301 (81), 299 (30), 287 (18), 269 (14), 241 (21), 227 (27). *Methyl ester 17*. [M – C₂H₂O]⁺ at *m/z* 392.2542, C₂₃H₃₆O₅ requires 392.2562; ¹H NMR: δ 0.77 and 1.03 (each 3H, s), 2.01 and 2.06 (each 3H, s), 2.54 (1H, br s, H-13), 2.83 (1H, dt, *J*₁ = 11, *J*₂ = *J*₃ = 7 Hz), 3.55 and 3.80 (each 1H, d, *J* = 11 Hz, H-18), 3.65 (3H, s), 4.66 (1H, m, H-7); EIMS *m/z* (rel. int.): 403 [M – MeO]⁺ (0.5), 392 (2), 374 (3), 359 (2), 319 (22), 314 (89), 301 (100), 299 (26), 287 (14), 285 (8), 269 (10), 241 (17), 227 (22), 196 (150).

Hydrogenation of aldehyde 6. Compound **6** (30 mg) was dissolved in EtOH and hydrogenated over C/Pd (5%), at room temp. for 12 hr. In this way a mixture of the aldehydes **12** and **13** was obtained. Oxidation with air of this mixture gave the corresponding acids. Methylation with CH₂N₂ and chromatography of the residue afforded the compound **16** and **17**, identical with the substances obtained above.

SeO₂ oxidation of candicandiol (9). Compound **9** (50 mg) was treated with SeO₂ (15 mg) in dioxan (5 ml) and H₂O (1.5 ml) at room temp. for 4 hr. Usual work-up and chromatography of the residue gave **21**, identical with the natural compound (canditriol).

Reduction of compound 5. Compound **5** (50 mg) was added to a soln of NaBH₄ (50 mg) and cerium trichloride (280 mg) in MeOH (7 ml). The mixture was stirred at room temp. for 90 min, diluted with H₂O and extracted as usual. Evaporation of the solvent and chromatography of the residue afforded starting material (**5**) (17 mg) and the triol **7** (28 mg). Acetylation of **7** in the usual way

afforded a compound identical with the acetylated natural product **8** (sinferol triacetate).

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